



Total synthesis of phytoprostane F₁ and its 16 epimer

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Abstract—The first synthesis of the two enantiomers of phytoprostane F₁ methyl ester **1** and **2** is described using the *syn-anti-syn* alcoxy ester **3** as starting material. © 2003 Elsevier Science Ltd. All rights reserved.

Isoprostanes (IsoPs) are a complex family of compounds produced, *in vivo*, from peroxidation of polyunsaturated fatty acids (AA, DHA, EPA) via a free-radical-catalyzed mechanism.¹ Thus, quantification of these IsoPs in the biological fluids and tissues as markers of lipid peroxidation appears as an important insight in the exploration of the role of the oxidative stress in human pathology.² Higher plants generally do not synthesize the precursor arachidonate required for isoPs formation, but rather utilize α -linolenic acid for the formation of isoprostane F₂-like compounds which have been termed phytoprostanes F₁.³ Jasmonates are established plant signal compounds that induce defense response.⁴ Preliminary data indicates that phytoprostanes also induce phytoalexins in a variety of plant species suggesting a possible function of phytoprostanes as mediators of plant defense reactions in response to oxidative stress.⁵ During the last two years, Zaroni et al. reported the first synthesis of phytoprostane A₁⁶ and Spur et al. just published the synthesis of 15-E₂-isoP (PPE₂).⁷

In connection with our program directed towards the synthesis of isoprostanes,⁸ we now report the first synthesis of *ent*-phytoprostane F₁ type I (PPF₁) **1** and its

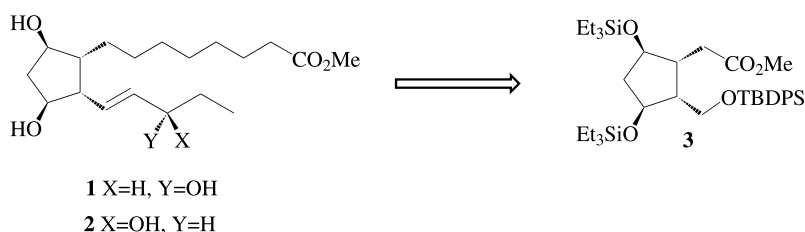
16-epimer **2** from the *syn-anti-syn* alcoxyester **3**⁹ (Scheme 1).

The synthesis of PPF₁ type I **1** and its 16(*R*) epimer **2** from the commercially available diacetone-D-glucose as starting material, is shown in Schemes 3 and 4. The first nine steps leading to cyclopentane alcoxyester **3** were achieved in 27% overall yield by using iodo pathway, according to our procedure.⁹

Synthon **4** was selected and prepared using the following procedure outlined in Scheme 2.

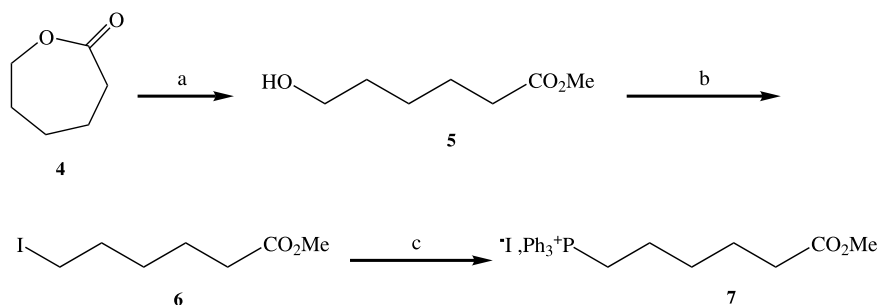
The commercially available ϵ -caprolactone was transformed to the corresponding hydroxy methyl ester **5** using acidic conditions in 97% yield. Halogenation with PPh₃/I₂ in CH₂Cl₂ gave iodo ester **6** in good yield. Phosphorylation of the iodide derivative **6** in the presence of PPh₃ and 1% of K₂CO₃ in the reaction mixture afforded the phophonium salt **7** in 98% yield.

The alcoxyester **3** was converted into the aldehyde **8** by treatment with DIBAL-H in anhydrous toluene with 97% yield (Scheme 3). The introduction of the α -chain of the phytoprostane was achieved by using the

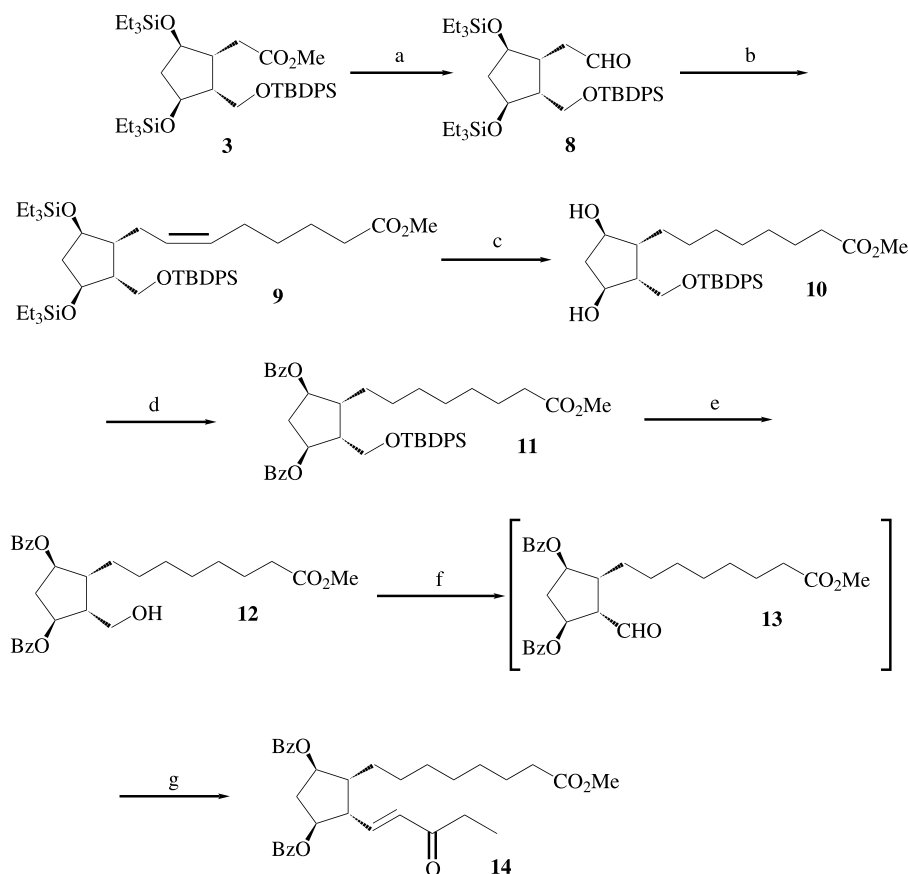


Scheme 1.

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Scheme 2. Reagents and conditions: (a) MeOH, H₂SO₄, rt, 30 min, 97%; (b) I₂, PPh₃, imidazole, CH₂Cl₂, –10°C, 2 h, 61%; (c) PPh₃, CH₃CN, 1% K₂CO₃, 18 h, 98%.



Scheme 3. Reagents and conditions: (a) 1.1 equiv. DIBAL-H (1 M in toluene), toluene, –80°C, 30 min, 97%; (b) 2.25 equiv. methoxycarbonylpentyltriphenylphosphonium iodide, 2.2 equiv. KN(SiMe₃)₂, THF, –80 to 20°C, 2 h, 70%; (c) H₂, Pd/C 10%, EtOH, 4 h, 98%; (d) 3 equiv. BzCl, pyridine, 20°C, 1 h, 77%; (e) HCl 3% in MeOH, 20°C, overnight, 80%; (f) periodinane, CH₂Cl₂, rt, 2 h; (g) 3.3 equiv. dimethyl 2-oxobutylphosphonate, 3 equiv. NaN(SiMe₃)₂, THF, 20°C, 30 min, 60%.

methoxycarbonylpentyltriphenyl phosphonium iodide **7**. The aldehyde **8** reacted with the ylide derived from this phosphonium salt and potassium hexamethyldisilyl amide as a base, in anhydrous THF at –80°C, to afford the pure (*Z*) enic ether **9** in 70% yield. No trace of *trans* compound could be detected by ¹³C and ¹H NMR analysis.

The *cis* double bond of **9** was reduced with H₂ on 10% Pd/C to give diol methyl ester **10** in 98% yield. Interestingly, during this hydrogenation we could avoid the deprotection of the triethylsilyl ethers. The protection of the hydroxy functions of **10** with benzoyl chlo-

ride in dry pyridine gave the colorless diester **11** in 77% yield.

The *tert*-butyldiphenylsilyl ether **11** was converted into the alcohol **12** with a solution of 3% hydrogen chloride¹⁰ in methanol/diethyl ether (1:1, v/v), prepared freshly from acetyl chloride and methanol, a method which proved to be much milder and to give a higher yield (80%) than TBAF in THF. Dess–Martin oxidation¹¹ of **12** in CH₂Cl₂ gave the unstable aldehyde **13** which was immediately used in the next step without purification. The condensation of **13** with dimethyl 2-oxobutylphosphonate,¹² in the presence of sodium

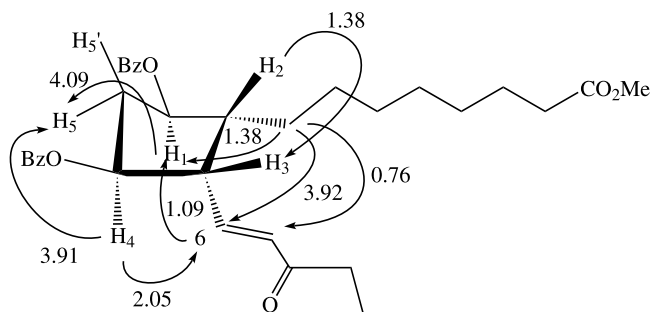


Figure 1. Observed NOEs resulting from irradiation of main protons are indicated with solid.

hexamethyldisilyl amide, in anhydrous THF at room temperature, afforded the *trans*- α,β enone ester **14** in 60% overall yield from the alcohol **12** (Scheme 3). We confirmed the relative configurations of the chiral centers by homonuclear ^1H steady-state-difference NOE spectroscopy (DNOES) experiments, as shown in Figure 1.

The diastereoselective reduction of the C-16 keto group in **14** with the chiral reducing agent¹³ (*S*)-BINAL-H proceeded smoothly and gave the desired pure 16(*S*) derivative **15** in 85% yield. Similarly, reduction of **14** using the (*R*)-BINAL-H gave the 16(*R*) epimer **16** in 87% yield (Scheme 4).

Finally, deprotection of the benzoyl groups in **15** and **16**, with 1N NaOH at room temperature, followed by excess of CH_2N_2 , afforded the desired *ent*-PPF₁ type I **2**¹⁴ and its 16 epimer **1**¹⁵ in 91 and 93% yields, respectively.

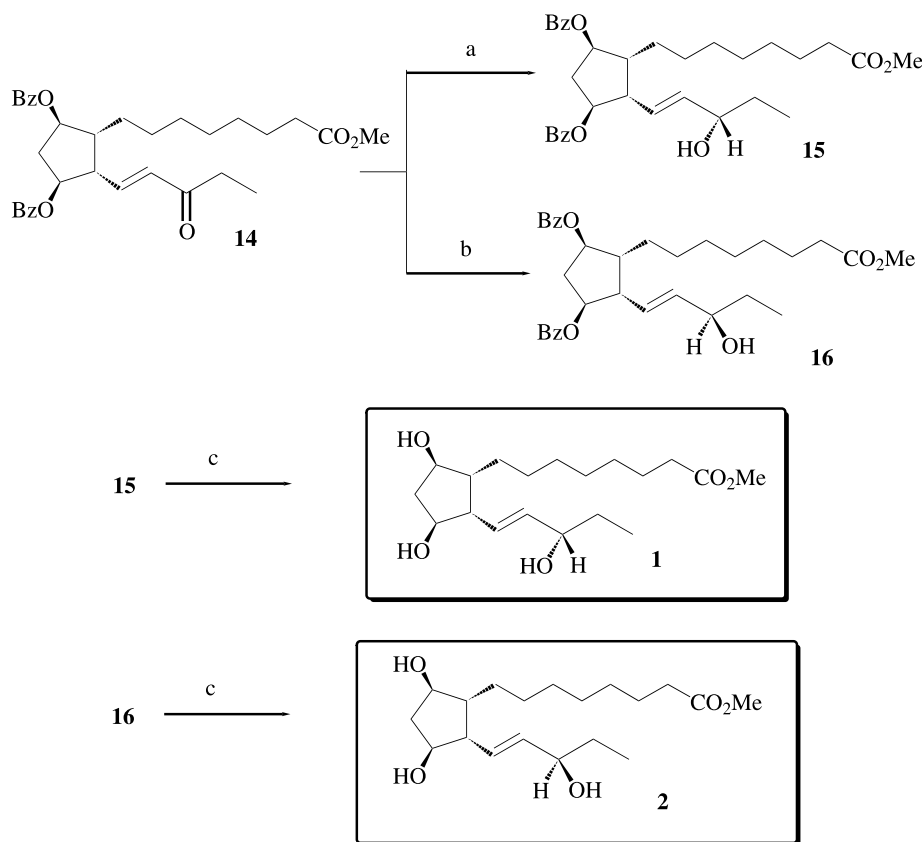
In conclusion, we have developed a practical synthesis of phytoprostane F₁ type I **2** and its 16 epimer **1** from our intermediate, the *syn-anti-syn* alcoxyester **3** in nine steps. The extension of this strategy towards type II PPF₁ and other phytoprostanes will be reported in due course.

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Scheme 4. Reagents and conditions: (a) (*S*)-BINAL-H, -100°C , 2 h, 85%; (b) (*R*)-BINAL-H, -100°C , 2 h, 87%; (c) 1N NaOH, THF–MeOH, rt, 1 h, then CH_2N_2 , 93%.

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14. Compound **2**: UV (ethanol) λ_{max} : 203 nm; ^1H NMR (400 MHz, CDCl_3): δ ppm 5.54 (dd, $J_{15,16}=7.1$ Hz, $J_{15,14}=15.2$ Hz, 1H, H-15), 5.40 (dd, $J_{14,13}=9.7$ Hz, $J_{14,15}=15.2$ Hz, 1H, H-14), 4.02–3.93 (m, 1H, H-16), 4.03–3.95 (m, 1H, H-12), 3.97–3.90 (m, 1H, H-10), 3.66 (s, 3H, OCH_3), 2.77–2.71 (m, 1H, H-13), 2.42 (dt, $J_{11,10}=J_{11,12}=6.8$ Hz, $J_{11,11'}=14.5$ Hz, 1H, H-11), 2.29 (t, $J_{2,3}=7.5$ Hz, 2H, H-2), 2.11–2.01 (m, 1H, H-9), 1.68–1.58 (m, 1H, H-11'), 1.66–1.55 (m, 2H, H-3), 1.65–1.45 (m, 2H, H-17), 1.33–1.25 (m, 2H, H-4), 1.33–1.19 (m, 8H, H-5, H-6, H-7 and H-8), 0.90 (t, $J_{18,19}=7.5$ Hz, 3H, H-18); ^{13}C NMR (100 MHz, CDCl_3): δ ppm 174.3 (C-1), 135.5 (C-15), 128.8 (C-14), 76.6 (C-10), 76.4 (C-12), 74.4 (C-16), 53.6 (C-13), 51.4 (OCH_3), 50.4 (C-9), 42.5 (C-11), 34.0 (C-2), 30.1 (C-17), 29.5 (C-8), 29.0 (C-4, C-5 and C-7), 28.0 (C-6), 24.8 (C-3), 9.6 (C-18); IR (NaCl) ν : 3520, 1720 cm^{-1} ; $[\alpha]_{\text{D}}^{20}=-30$ (c 1×10^{-2} , MeOH).
15. Compound **1**: UV (ethanol) λ_{max} : 203 nm; ^1H NMR (400 MHz, CDCl_3): δ ppm 5.57 (dd, $J_{15,16}=6.2$ Hz, $J_{15,14}=15.4$ Hz, 1H, H-15), 5.43 (dd, $J_{14,13}=5.4$ Hz, $J_{14,15}=15.4$ Hz, 1H, H-14), 4.05–3.96 (m, 1H, H-16), 4.04–3.99 (m, 1H, H-12), 3.98–3.93 (m, 1H, H-10), 3.66 (s, 3H, OCH_3), 2.79–2.73 (m, 1H, H-13), 2.41 (dt, $J_{11,10}=J_{11,12}=6.8$ Hz, $J_{11,11'}=14.5$ Hz, 1H, H-11), 2.29 (t, $J_{2,3}=7.7$ Hz, 2H, H-2), 2.12–2.04 (m, 1H, H-9), 1.68–1.59 (m, 1H, H-11'), 1.66–1.55 (m, 2H, H-3), 1.60–1.48 (m, 2H, H-17), 1.33–1.25 (m, 2H, H-4), 1.33–1.18 (m, 8H, H-5, H-6, H-7 and H-8), 0.90 (t, $J_{18,19}=7.3$ Hz, 3H, H-18); ^{13}C NMR (100 MHz, CDCl_3): δ ppm 174.3 (C-1), 135.3 (C-15), 128.7 (C-14), 76.8 (C-10), 76.5 (C-12), 73.8 (C-16), 53.4 (C-13), 51.4 (OCH_3), 50.3 (C-9), 42.5 (C-11), 34.0 (C-2), 30.1 (C-17), 29.4 (C-8), 29.0 (C-4, C-5 and C-7), 28.0 (C-6), 24.8 (C-3), 9.6 (C-18); IR (NaCl) ν : 3520, 1720 cm^{-1} ; $[\alpha]_{\text{D}}^{20}=-13$ (c 1×10^{-2} , MeOH).